

Neuronal failure in Alzheimer's disease: a view through the oxidative stress looking-glass

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Considerable debate and controversy surround the cause(s) of Alzheimer's disease (AD). To date, several theories have gained notoriety, however none is universally accepted. In this review, we provide evidence for the oxidative stress-induced AD cascade that posits aged mitochondria as the critical origin of neurodegeneration in AD.

Keywords: Alzheimer's disease; amyloid-beta; free radicals; mitochondria; mitochondrial dynamics; oxidative stress

Introduction

Alzheimer's disease (AD) is a neurodegenerative disorder responsible for the cognitive deterioration of 26.6 million people worldwide. The prevalence is projected to increase by ~3 fold to 106.8 million by the year 2050, with 1 in 86 people living with AD and 59% of the cases in Asia^[1]. Age is the primary risk factor for AD such that its incidence is 15% among individuals >65 years old and reaches ~50% among those >85^[2]. While many specific aspects of AD have been documented, there has yet to be any clear understanding of its pathological initiation and progression. Molecular aberrations in the cell elicit neuronal failure through various well-established mechanisms, including oxidative stress^[3], abnormal protein folding and aggregation^[4], cell cycle dysregulation^[5], mitochondrial dysfunction^[6], synaptic failure^[7, 8], inflammation^[9], loss of calcium regulation^[10], defective cholesterol metabolism^[11, 12], vascular alterations^[13, 14], and neurotrophin deprivation^[11]. However, the causal relationships governing these factors remain unknown.

AD is characterized by the successive degeneration of neurons first in the entorhinal cortex of the mediotemporal

lobe, followed by the CA1 region of the hippocampus, CA2/3, CA4, and the neocortex^[15]. Despite this predictable vulnerability, the molecular mechanisms governing it are unclear.

Amyloid- β (A β) and the hyperphosphorylated form of the microtubule-associated protein tau are hallmarks of AD pathology, and are critically involved in neuronal death. While the function of A β is unclear, its accumulation into insoluble fibrils and plaques increases over the course of AD progression, causing a severe cellular burden. The A β peptide is formed by the proteolytic cleavage of its protein precursor, amyloid- β protein precursor (A β PP), via the sequential actions of β -site A β PP-cleaving enzyme 1 and γ -secretase, a protein complex with presenilins 1 and 2 at its catalytic core^[11, 16]. A β is associated with multiple cascades that are thought to result in neuronal damage^[17, 18] and as such, it is generally considered to be the primary mediator of neurodegeneration in AD^[19]. However, the lack of therapeutic translation has become a major criticism of the amyloid cascade hypothesis^[20].

The microtubule-associated protein tau is similarly associated with AD progression, and its accumulation in the form of neurofibrillary tangles (NFTs) strongly correlates

with dementia in AD^[11, 21]. NFTs are used as a post-mortem diagnostic criterion for AD^[22]. Tau is phosphorylated at serine and threonine residues that flank the microtubule-binding domain. Upon hyperphosphorylation, tau is unable to bind and stabilize cytoskeletal microtubules; instead it self-aggregates into NFTs in and around the cell^[23].

AD is typically sporadic; nearly 90% of all cases are sporadic in nature and characterized as late-onset AD (LOAD), with an age of onset of ~65 years. Familial AD (FAD), on the other hand, is an early-onset, genetic condition caused by several known autosomal dominant mutations; its age of onset is typically ~40–60 years^[24]. Mutations that yield FAD all involve or result in an alteration in the ratio of A β ₄₂ to A β ₄₀; this is perhaps the most touted evidence both for and against^[25, 26] the amyloid cascade hypothesis^[27]. Dominantly-inherited mutations in presenilin 1 (PS1), an essential component of the γ -secretase complex, account for the majority of FAD cases, and >170 such mutations in PS1 have been identified on chromosome 14^[28]. In addition, mutations in presenilin 2 (PS2) (on chromosome 1^[29]) and A β PP (on chromosome 21^[30]) also elicit FAD, although they are less prevalent. These mutations also affect the A β _{42/40} ratio. Importantly, both LOAD and FAD present identical brain lesions and patterns of neurodegeneration. Notably and somewhat contradictory, LOAD leads to a reduced A β _{42/40} ratio, whereas FAD leads to an increase^[31].

Neuronal Oxidative Stress: Sources and Vulnerabilities

Aerobic respiration, like any other biochemical or physical process, is not 100% efficient. The mechanism by which mitochondria process carbohydrates and establish a proton gradient for the synthesis of ATP (i.e., the tri-carboxylic acid (TCA) cycle and oxidative phosphorylation) involves the controlled transfer of electrons from strong reducing agents to strong oxidizing agents. Some of the free radicals thus generated escape; rather than appropriately transporting their electron to the next molecule in the cascade, they abandon the inner membrane of the mitochondrion and impose alterations on other macromolecules within the cell. These alterations are often detrimental: DNA/RNA oxidation may yield fragmentation and deficiencies in repair machinery^[32, 33]; oxidative modification of enzymes

and metabolic signaling proteins may lead to metabolic impairments^[34]; and protein cross-linkages and an impaired proteolysis network resulting from oxidative modification may render proteins insoluble and prone to abnormal aggregation^[35–37].

The ability of the cell to adapt to oxidative damage is robust, yet finite. Endogenous antioxidants are intended to sequester the formation of free radicals. Superoxide dismutase, for instance, catalyzes the conversion of the potent free radical superoxide to hydrogen peroxide and water. Other important oxidative stress-handling proteins include catalase, glutathione peroxidase, glutathione reductase, and heme oxygenase^[38]. Under normal conditions, the majority of free radicals are sequestered within the mitochondria (i.e., their place of origin). Notably, however, three important factors (age, metabolic demand, and disease) exacerbate the vulnerability of cells to oxidative burden.

Statistically, the longer a cell is respiring, the more reactive oxidative species (ROS) go unsequestered within that cell. It is estimated that cells use 10¹³ molecules of oxygen on average per day, with 1% of the associated reactions producing unsequestered ROS. Therefore, 10¹¹ molecules of ROS are generated in a given cell every day. Even with efficient antioxidant mechanisms, some ROS unavoidably escape and cause damage to the surroundings that may accumulate with time, especially in long-living cells. Age is thus a great risk factor for oxidative stress. Likewise, high metabolic activity, which requires more oxidative phosphorylation per unit time and thus involves a greater number of toxic species generated per unit time, also renders a cell more vulnerable to oxidative insults. In particular, the neuron, which requires energy for axonal transport, vesicular release, ion pump operation, electrochemical gradient maintenance, and the like, requires much more oxygen per unit time than any other cell type in the body. Metabolic demand is therefore another risk factor for oxidative stress. Disease conditions that produce mutations in cellular antioxidant defensive machinery provide a third risk factor for oxidative damage.

Altogether, the brain is tremendously vulnerable to oxidative damage. Although it constitutes only 2–3% of total body mass, 20% of the basal oxygen supply is used by the brain. Moreover, neurons are among the longest-living cells in the body and need the same metabolic

machinery for continuation of survival and functioning for decades. The presence of transition metals in the brain that catalyze oxidative reactions further increases the appearance of oxidative pathologies. Iron and copper, for instance, increase the likelihood of an oxidation/reduction reaction^[39], and increases in the regional concentrations of these metals in brain compound the risk factors described above. Furthermore, the brain suffers a relative paucity of antioxidant systems as well as an increase in polyunsaturated fatty acids (prime targets for ROS)^[40]. So an aging brain is tremendously susceptible to oxidative deterioration.

Oxidative Stress: a Prominent and Early Feature of Alzheimer's Disease

Free radical-induced damage to macromolecules has been well documented in AD. Deposition of redox-active ions, which are capable of generating the most damaging hydroxyl radicals through the Fenton reaction, is likely to exacerbate both the spectrum of molecules and the cellular areas affected. DNA and RNA oxidation, marked by increased levels of 8-hydroxy-2-deoxyguanosine and 8-hydroxyguanosine (8-OHG), suggest a higher frequency of DNA fragmentation and aberration in DNA repair^[41]. Oxidative modification of metabolic proteins, such as creatine kinase BB, cytochrome c oxidase (COX), and ketoglutarate dehydrogenase complex (KGDH), has been demonstrated by elevated levels of protein carbonyl and nitration of tyrosine residues^[42] and indicates impaired metabolic activity. Lipid peroxidation, marked by thiobarbituric acid reactive substances, malondialdehydes, 4-hydroxy-2-transnonenal (HNE), and isoprostane, as well as by altered phospholipid composition, suggests altered membrane integrity. Oxidative modification of sugars, marked by increased glycation and glycooxidation, indicates an impaired cellular ability to adequately process critical carbohydrates^[43]. Each of these aspects is elevated in AD compared to control cases^[42].

Moreover, mitochondrial abnormalities, attributed to oxidative stress-related damage, are well-established in AD. As stated above, specific enzymes involved in electron transport and the TCA cycle, such as KGDH and COX, are modified *via* ROS in AD^[44,45]. Mitochondrial DNA (mtDNA) modification^[46] and calcium dysregulation^[47-49] occur

in AD to a great extent, and increased numbers of mitochondria with broken cristae, altered size and shape, and aberrant intracellular localization are elevated in AD neurons^[47, 50, 51]. These latter phenomena are the result of impaired mitochondrial dynamics that can be caused by and also amplify oxidative stress^[52]. Mitochondrial fission and fusion, the ongoing processes that ensure organelle stability within the dynamic cellular environment, rely on critical membrane proteins. Fusion enables the exchange of lipid membrane in inter-mitochondrial components (i.e., mtDNA and fission/fusion proteins) such that the cell is populated by healthy, normal mitochondria^[53]. Fission, on the other hand, coupled with fusion and autophagy, enables the sequestration and elimination of irreversibly damaged mitochondria and mitochondrial content^[54]. These dynamics are the necessary means by which the cell dampens its inevitable accumulation of oxidative free radical-induced damage. Over the lifetime of a cell, however, oxidative damage and transcriptional errors (due to alteration of mtDNA) reach a critical threshold. Oxidative stress ultimately propagates throughout the cell as mitochondria become abnormally shaped and localized^[47, 55-58].

Importantly, increasing evidence demonstrates that the oxidative modifications in vulnerable neurons in AD occur prior to the hallmark pathologies of the disease^[35]. That is, the markers of oxidative damage are found in the cytoplasm of neurons prior to any indication of degeneration in AD brains^[59, 60]. In fact, the hallmark pathologies such as amyloid plaques and NFTs may be an adaptation in response to elevated oxidative stress. More recent studies showed that patients who suffer from mild cognitive impairment (MCI), considered to be a prodromal stage of AD, are depleted in antioxidants along with increased lipid peroxidation in the plasma and lymphocytes^[61]. Elevated protein/RNA oxidation and redox-active iron are also documented in the brains of MCI patients^[32,62,63]. Transgenic mouse models of AD also demonstrate that oxidative damage precedes A β deposition^[64]. The mitochondrial abnormalities described above provide considerable support for this primacy. Specifically, vulnerable neurons in AD and MCI exhibit severe metabolic deficiencies before any clinical manifestations of disease. Neuroimaging studies and neuropsychological tests have shown impaired cerebral metabolism prior to any evidence of functional impairment or brain atrophy induced by AD

pathologies^[51, 65]. This metabolic impairment is likely the result of the alterations in mitochondrial dynamics and intracellular localization^[66], which, in turn, increases the production of oxidative damage to mtDNA and membrane proteins leading to a vicious cycle.

Alzheimer-Specific Factors and Oxidative Stress

Besides the fact that oxidative stress characterizes aging, genetic and pathological factors associated with AD are also implicated in the regulation of oxidative stress. For example, presenilins are needed for the import of ~50% of cellular copper and zinc^[67]. Increased presenilin 2 expression increases DNA fragmentation and induces apoptotic changes^[68], which are important consequences of oxidative damage. The translation of APP is regulated by iron influx *via* an iron-responsive element RNA stem loop in its 5'-untranslated region^[69]. APP is the dedicated ferroxidase in neurons responsible for the export of iron, and the elevation of redox-active iron in neurons in AD is likely due to the reduced ferroxidase activity of APP inhibited by zinc^[70]. ApoE is a strong chelator of copper and iron, both of which are important redox-active transition metals^[68]. ApoE is beneficial against free radicals as it reduces neuronal death caused by hydrogen peroxide and A β through antioxidant activity in an isoform-dependent manner, the E4 isoform being the least effective^[71]. Interestingly, apoE is sensitive to attack by free radicals, and this sensitivity is also isoform-dependent, E4 being the most vulnerable^[72]. Indeed, cerebral cortex samples obtained at autopsy from patients with APP or presenilin gene mutations or the apoE4 allele demonstrate higher oxidative stress^[73, 74].

A β itself may cause oxidative damage in surrounding cells through, for example, activation of microglia and astrocytes that activate multiple ROS pathways^[75], including upregulating L-tryptophan metabolism through the kynurenine pathway^[76]. Several intermediates in this pathway, namely 3-hydroxykynurenine and quinolinic acid, are neurotoxic, and the A β -induced increase in their production subjects surrounding neurons to oxidative damage^[77, 78]. A β -induced lipid peroxidation yields several reactive aldehydes (including HNE) that can harmfully modify membrane proteins. HNE production specifically disrupts iron homeostasis by impairing glucose

transporters^[79–82]. Iron and copper accumulation within neuritic plaques (mainly composed of A β) further facilitates free-radical generation^[83]. When A β binds catalytic amounts of copper, hydrogen peroxide is generated, which contributes to oxidative stress as it produces hydroxyl radicals *via* the Fenton reaction.

A β has antioxidant effects intracellularly and may actually be secreted as a compensatory measure against oxidative stress^[35, 84]. *In vivo* and *in vitro* studies of neuronal responses to oxidative stress indicate an increase in A β production^[85], which is followed by a corresponding reduction in oxidative stress^[86, 87]. Moreover, markers of oxidative stress, such as 8-OHG, are reduced in neuronal populations characterized by A β deposition, and elevated in vulnerable regions lacking A β ^[86, 87]. The data suggest that neurons may be salvaged from oxidative stress by A β .

Tau hyperphosphorylation is a pathological result of oxidative stress^[59, 84, 87–89]. Interestingly, most neuronal loss during the course of neurodegeneration occurs where the levels of oxidative stress are highest, and the subsequent deposition of NFTs decreases these levels^[59]. Of note, neurons with accumulated NFTs are able to survive for decades and become functionally integrated in cortical circuits^[90, 91]. Phosphorylated tau antagonizes apoptosis by stabilizing beta-catenin^[92]. Regardless, the abnormal accumulation of hyperphosphorylated tau in the form of NFTs occurs subsequent to oxidative stress-induced damage.

Oxidative Stress and Alzheimer's Disease: Ubiquity versus Specificity

Disease is defined by deficits in specific functional output, with the underlying anatomical and biochemical/structural changes elicited by insults and adaptations and/or failure of adaptations. Interestingly, oxidative stress is a prominent biochemical phenomenon not only in AD, but in almost all of the major neurodegenerative diseases, including Parkinson disease, amyotrophic lateral sclerosis, and Huntington disease^[93]. Since oxidative stress contributes largely to aging, and age is the greatest risk factor for all neurodegenerative diseases, it is not surprising that oxidative stress is involved in pathogenesis of each of these diseases. However, given the different neuronal populations and distinct pathologies that characterize each

disease, the ubiquity of oxidative stress raises a question about the specificity of its involvement. In AD, for example, elevated oxidative stress (and neuronal damage) occurs in areas such as hippocampus and cortex, but not in the cerebellum, midbrain, or pons^[42]. Similarly in Parkinson disease, enhanced oxidative stress primarily localizes to the substantia nigra and the basal ganglia^[94]. If oxidative stress is indeed a fundamental pathologic process in these diseases, the region-specific neuronal deterioration demands explanation.

Although further study is necessary, it is likely that the specific characteristics of the neuronal populations involved contribute to such selectivity. Neurons with long axons and multiple synapses have higher metabolic demands that may render them more prone to oxidative attack at a steady-state level and thus become more vulnerable to additional disease-related changes. For example, CA1 neurons have higher levels of superoxide anion than parietal cortical or CA3 neurons, which is at least one of the reasons why they are more vulnerable to global ischemia-induced cell death^[95]. Dopaminergic neurons are also exposed to high steady-state levels of oxidative stress produced by the metabolism of dopamine, which make them more vulnerable to insults that affect dopamine metabolism^[96].

The particular vulnerability of a given individual to developing cumulative oxidative damage to the hippocampus (and thus suffering from AD) rather than the basal ganglia probably reflects a particular arrangement of predisposing genetic and environmental factors. Such initiating factors may not be a single event, but more likely a complex interaction between individual genetic/epigenetic backgrounds and the environment that ultimately determines the specific neuronal population affected. Overall, different diseases may display distinct oxidative stress patterns that likely will lead to a different homeostatic balance that is finely tuned to minimize cell death. More studies are needed to understand the role of oxidative stress in each disease.

The Age-Related Mitochondrial Cascade in Alzheimer's Disease

The evidence listed above reveals the following: (1) AD is primarily an age-related disorder; (2) the brain is the most metabolically demanding organ in the body; (3) free

radical release invariably results from the tremendous amount of oxidation/reduction reactions within neurons; (4) resident antioxidant systems, though robust, are finite in capacity; (5) mitochondria are the metabolic center of the cell and experience the earliest detriments in AD; and (6) A β peptides and NFTs accumulate after oxidative stress has already incurred severe damage, and seem to be a compensatory response to cerebral oxidative stress.

Fitting the pieces together, we propose that over the course of a lifetime, statistically inevitable free-radical damage accumulates within mitochondria, affecting mtDNA, membrane proteins, and calcium homeostasis. The antioxidant response of neurons is able to hold back the effects of this stress for years or decades; metabolic deficits resulting from mitochondrial dysfunctions and perturbed localizations do not appear until adult life, and are the very first aspect of dementia. However, the increasing levels of transition metals and polyunsaturated fatty acids in the brain render neurons particularly susceptible to damage, and once mitochondrial mutations become widespread within the cell, compensatory responses, such as A β secretion, are initiated to defend against further damage. Eventually, secreted A β becomes subject to oxidative stress (i.e., dityrosine cross-linkages inhibit its solubility, promoting aggregation^[62, 97]). The cascade worsens as A β induces neuronal defects, producing further oxidative stress, inflammation, and synaptic dysfunction. The well-established pathophysiological inclusions of AD, including cell cycle aberration, inflammation, synaptic dysfunction, and loss of calcium regulation, become increasingly relevant, and affected neurons ultimately die. Thus, after decades of stress and increasing malfunction, cognitive decline spreads and yields clinical AD (Fig. 1).

Though its logic is appealing, this description of AD, like all previous hypotheses posited as explaining the disease, suffers significant shortcomings. First, it does not adequately explain the predictable and consistent spread of neurodegeneration from the entorhinal cortex of the mediotemporal lobe, through the hippocampus, and into the neocortex. Perhaps this orderly passage reflects the metabolic demands of distinct regions within the brain; the entorhinal cortex may require the highest metabolic activity due to its involvement in memory formation and retrieval (an ongoing process throughout life). It is thus the first to be affected by oxidative stress accumulation that

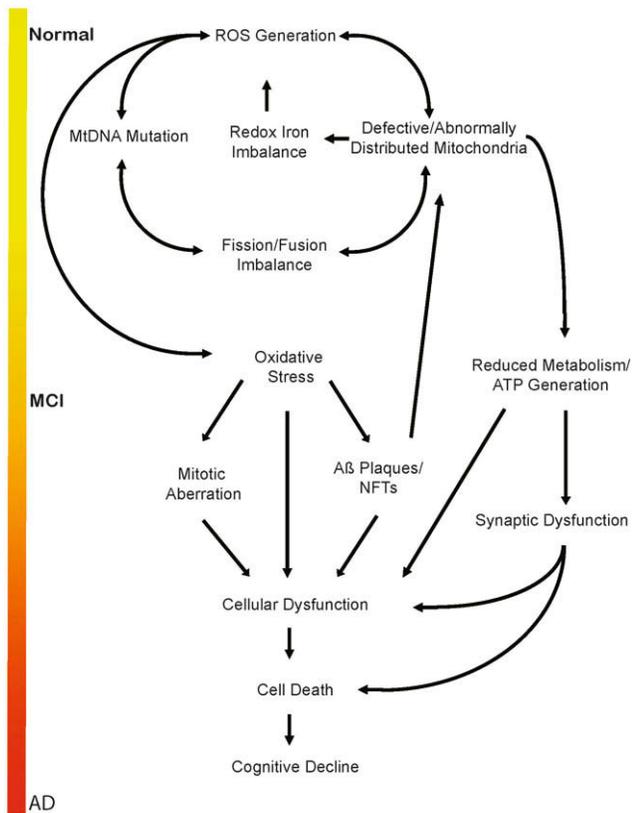


Fig. 1. Oxidative stress-induced cascade of Alzheimer disease (AD). The inevitable generation of free radicals within neurons eventually compromises the integrity of mitochondria, leading to a cascade of destructive events that elicits the hallmark features of AD. After years of accumulation, neuronal death and cognitive decline occur. MCI, mild cognitive impairment; NFTs, neurofibrillary tangles; ROS, reactive oxidative species.

leads to degeneration by a matter of simple probability. The remaining neurons in the pathway would suffer by connective association, and the disease would spread. Notably, this postulate lacks evidence, and its potential validity merits further investigation.

The mitochondrial cascade also seems to fail to explain the critical role of AD mutations in FAD and Down syndrome. A more careful look, however, ameliorates this misperception. In particular, the timeline of LOAD is such that A β accumulation and aggregation occurs late in adult life, causing deficits in cognition typically after age 65. The mitochondrial cascade attributes this latency to the prolonged period of oxidative stress accumulation, which takes years to have any metabolic deficits and subsequent

A β secretion. Once secreted, A β , however, is oxidatively processed into insoluble fibrils, and damages surrounding cells *via* a variety of mechanisms. FAD represents a shortcut in this otherwise lengthy process. Mutations that increase the cleavage and processing of A β PP subject neurons to the early secretion and aggregation of A β . Thus, affected patients skip the necessary accumulation of mitochondrial damage, and with aberrantly produced A β , display clinical symptoms much earlier than LOAD patients, regardless of the endogenous oxidative environment. Still, little experimental evidence supports this postulate, but it seems quite likely given the mountains of data.

Therapeutic Implications and Conclusions

We are currently faced with a strong incentive to produce a therapeutic agent that prevents AD. Based on the evidence presented here, the most likely method of adequate prevention may come from antioxidation. Preventing the accumulation of damaging species within vulnerable neurons would ideally protect them from the deleterious cascades that ROS accumulation yields. Unfortunately, antioxidant therapies have had little success thus far^[98]. It should be emphasized that the failure of antioxidant clinical trials does not necessarily nullify the contribution of oxidative stress to the disease; other factors such as patient selection (it would be preferable to select patients with low antioxidants) and monitoring/analysis of the responders (it would be preferable to monitor oxidative stress surrogate markers before and during the treatment to make certain of compliance) are important for clinical studies. Better antioxidants, especially those that target the major ROS production sites, are needed. Several agents under investigation, such as MitoQ, acetyl-L-carnitine, and α -lipoic acid, do show potential; however, an effective method of treatment is far from complete^[47, 99-110]. Based on the many prospective studies and the complexity of the redox system *in vivo*, a balanced combination of several antioxidants may also be needed to have a significant effect on the prevention of AD, but it appears that not much has been done on this aspect. Although agents that ameliorate pathologies secondary to AD are under increasing scrutiny^[22], these treatments would only slow the course of disease progression and could not achieve its full eradication. To do so, the community must acknowledge all

aspects of disease pathogenesis and establish an unbiased understanding of its progression. Only then can research be adequately focused such that appropriate therapeutic intervention is possible. We here pose the oxidative stress “looking glass” as our best hope.

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REFERENCES

- [1] Brookmeyer R, Johnson E, Ziegler-Graham K, Arrighi HM. Forecasting the global burden of Alzheimer's disease. *Alzheimers Dement* 2007, 3: 186–191.
- [2] Smith MA. Alzheimer disease. *Int Rev Neurobiol* 1998, 42: 1–54.
- [3] Yan MH, Wang X, Zhu X. Mitochondrial defects and oxidative stress in Alzheimer disease and Parkinson disease. *Free Radic Biol Med* 2013, 62: 90–101.
- [4] Walker LC, Diamond MI, Duff KE, Hyman BT. Mechanisms of protein seeding in neurodegenerative diseases. *JAMA Neurol* 2013, 70: 304–310.
- [5] Moh C, Kubiak JZ, Bajic VP, Zhu X, Smith MA, Lee HG. Cell cycle deregulation in the neurons of Alzheimer's disease. *Results Probl Cell Differ* 2011, 53: 565–576.
- [6] Zhu X, Perry G, Smith MA, Wang X. Abnormal mitochondrial dynamics in the pathogenesis of Alzheimer's disease. *J Alzheimers Dis* 2013, 33 Suppl 1: S253–262.
- [7] Selkoe DJ. Alzheimer's disease is a synaptic failure. *Science* 2002, 298: 789–791.
- [8] Sheng M, Sabatini BL, Sudhof TC. Synapses and Alzheimer's disease. *Cold Spring Harb Perspect Biol* 2012, 4.
- [9] Krstic D, Knuesel I. Deciphering the mechanism underlying late-onset Alzheimer disease. *Nat Rev Neurol* 2013, 9: 25–34.
- [10] Camandola S, Mattson MP. Aberrant subcellular neuronal calcium regulation in aging and Alzheimer's disease. *Biochim Biophys Acta* 2011, 1813: 965–973.
- [11] Querfurth HW, LaFerla FM. Alzheimer's disease. *N Engl J Med* 2010, 362: 329–344.
- [12] Kivipelto M, Helkala EL, Laakso MP, Hanninen T, Hallikainen M, Alhainen K, *et al.* Midlife vascular risk factors and Alzheimer's disease in later life: longitudinal, population based study. *BMJ* 2001, 322: 1447–1451.
- [13] de la Torre JC. Pathophysiology of neuronal energy crisis in Alzheimer's disease. *Neurodegener Dis* 2008, 5: 126–132.
- [14] Zhu X, Smith MA, Honda K, Aliev G, Moreira PI, Nunomura A, *et al.* Vascular oxidative stress in Alzheimer disease. *J Neurol Sci* 2007, 257: 240–246.
- [15] Schonheit B, Zarski R, Ohm TG. Spatial and temporal relationships between plaques and tangles in Alzheimer-pathology. *Neurobiol Aging* 2004, 25: 697–711.
- [16] Hebert LE, Scherr PA, Bienias JL, Bennett DA, Evans DA. Alzheimer disease in the US population: prevalence estimates using the 2000 census. *Arch Neurol* 2003, 60: 1119–1122.
- [17] Selkoe DJ. Alzheimer's disease: genes, proteins, and therapy. *Physiol Rev* 2001, 81: 741–766.
- [18] Tanzi RE, Bertram L. Twenty years of the Alzheimer's disease amyloid hypothesis: a genetic perspective. *Cell* 2005, 120: 545–555.
- [19] Selkoe DJ. Alzheimer's disease results from the cerebral accumulation and cytotoxicity of amyloid beta-protein. *J Alzheimers Dis* 2001, 3: 75–80.
- [20] Herrup K. Reimagining Alzheimer's disease—an age-based hypothesis. *J Neurosci* 2010, 30: 16755–16762.
- [21] Grundke-Iqbal I, Iqbal K, Tung YC, Quinlan M, Wisniewski HM, Binder LI. Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc Natl Acad Sci U S A* 1986, 83: 4913–4917.
- [22] Bonda DJ, Lee HP, Lee HG, Friedlich AL, Perry G, Zhu X, *et al.* Novel therapeutics for Alzheimer's disease: an update. *Curr Opin Drug Discov Devel* 2010, 13: 235–246.
- [23] Wang JZ, Xia YY, Grundke-Iqbal I, Iqbal K. Abnormal hyperphosphorylation of tau: sites, regulation, and molecular mechanism of neurofibrillary degeneration. *J Alzheimers Dis* 2013, 33 Suppl 1: S123–139.
- [24] Goedert M, Strittmatter WJ, Roses AD. Alzheimer's disease. Risky apolipoprotein in brain. *Nature* 1994, 372: 45–46.
- [25] Castellani RJ, Lee HG, Siedlak SL, Nunomura A, Hayashi T, Nakamura M, *et al.* Reexamining Alzheimer's disease: evidence for a protective role for amyloid-beta protein precursor and amyloid-beta. *J Alzheimers Dis* 2009, 18: 447–452.
- [26] Castellani RJ, Lee HG, Zhu X, Perry G, Smith MA. Alzheimer disease pathology as a host response. *J Neuropathol Exp Neurol* 2008, 67: 523–531.
- [27] Hardy J, Duff K, Hardy KG, Perez-Tur J, Hutton M. Genetic dissection of Alzheimer's disease and related dementias: amyloid and its relationship to tau. *Nat Neurosci* 1998, 1: 355–358.
- [28] Schellenberg GD, Bird TD, Wijsman EM, Orr HT, Anderson L, Nemens E, *et al.* Genetic linkage evidence for a familial Alzheimer's disease locus on chromosome 14. *Science* 1992, 258: 668–671.

- [29] Levy-Lahad E, Wijsman EM, Nemens E, Anderson L, Goddard KA, Weber JL, *et al.* A familial Alzheimer's disease locus on chromosome 1. *Science* 1995, 269: 970–973.
- [30] Goate A, Chartier-Harlin MC, Mullan M, Brown J, Crawford F, Fidani L, *et al.* Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature* 1991, 349: 704–706.
- [31] Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. *Circ Res* 2000, 87: 840–844.
- [32] Nunomura A, Tamaoki T, Motohashi N, Nakamura M, McKeel DW, Jr., Tabaton M, *et al.* The earliest stage of cognitive impairment in transition from normal aging to Alzheimer disease is marked by prominent RNA oxidation in vulnerable neurons. *J Neuropathol Exp Neurol* 2012, 71: 233–241.
- [33] Bradley-Whitman MA, Timmons MD, Beckett TL, Murphy MP, Lynn BC, Lovell MA. Nucleic Acid Oxidation: An early feature of Alzheimer's disease. *J Neurochem* 2014 128(2): 294–304.
- [34] Hardas SS, Sultana R, Clark AM, Beckett TL, Szweda LI, Murphy MP, *et al.* Oxidative modification of lipoic acid by HNE in Alzheimer disease brain. *Redox Biol* 2013, 1: 80–85.
- [35] Sayre LM, Smith MA, Perry G. Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Curr Med Chem* 2001, 8: 721–738.
- [36] Di Domenico F, Coccia R, Cocciolo A, Murphy MP, Cenini G, Head E, *et al.* Impairment of proteostasis network in Down syndrome prior to the development of Alzheimer's disease neuropathology: redox proteomics analysis of human brain. *Biochim Biophys Acta* 2013, 1832: 1249–1259.
- [37] Zhu X, Castellani RJ, Moreira PI, Aliev G, Shenk JC, Siedlak SL, *et al.* Hydroxynonenal-generated crosslinking fluorophore accumulation in Alzheimer disease reveals a dichotomy of protein turnover. *Free Radical Biology and Medicine* 2012, 52: 699–704.
- [38] Aksenov MY, Tucker HM, Nair P, Aksenova MV, Butterfield DA, Estus S, *et al.* The expression of key oxidative stress-handling genes in different brain regions in Alzheimer's disease. *J Mol Neurosci* 1998, 11: 151–164.
- [39] Sayre LM, Perry G, Smith MA. Redox metals and neurodegenerative disease. *Curr Opin Chem Biol* 1999, 3: 220–225.
- [40] Pratico D. Oxidative stress hypothesis in Alzheimer's disease: a reappraisal. *Trends Pharmacol Sci* 2008, 29: 609–615.
- [41] Nunomura A, Moreira PI, Castellani RJ, Lee HG, Zhu X, Smith MA, *et al.* Oxidative damage to RNA in aging and neurodegenerative disorders. *Neurotox Res* 2012, 22: 231–248.
- [42] Sultana R, Butterfield DA. Oxidative modification of brain proteins in Alzheimer's disease: perspective on future studies based on results of redox proteomics studies. *J Alzheimers Dis* 2013, 33 Suppl 1: S243–251.
- [43] Zhu X, Su B, Wang X, Smith MA, Perry G. Causes of oxidative stress in Alzheimer disease. *Cell Mol Life Sci* 2007, 64: 2202–2210.
- [44] Gibson GE, Shi Q. A mitocentric view of Alzheimer's disease suggests multi-faceted treatments. *J Alzheimers Dis* 2010, 20 Suppl 2: S591–607.
- [45] Schon EA, Przedborski S. Mitochondria: the next (neurode) generation. *Neuron* 2011, 70: 1033–1053.
- [46] Cheng X, Kanki T, Fukuoh A, Ohgaki K, Takeya R, Aoki Y, *et al.* PDIP38 associates with proteins constituting the mitochondrial DNA nucleoid. *J Biochem* 2005, 138: 673–678.
- [47] Hirai K, Aliev G, Nunomura A, Fujioka H, Russell RL, Atwood CS, *et al.* Mitochondrial abnormalities in Alzheimer's disease. *J Neurosci* 2001, 21: 3017–3023.
- [48] Keller JN, Guo Q, Holtsberg FW, Bruce-Keller AJ, Mattson MP. Increased sensitivity to mitochondrial toxin-induced apoptosis in neural cells expressing mutant presenilin-1 is linked to perturbed calcium homeostasis and enhanced oxyradical production. *J Neurosci* 1998, 18: 4439–4450.
- [49] Coskun PE, Beal MF, Wallace DC. Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proc Natl Acad Sci U S A* 2004, 101: 10726–10731.
- [50] Bonda DJ, Wang X, Perry G, Smith MA, Zhu X. Mitochondrial dynamics in Alzheimer's disease: opportunities for future treatment strategies. *Drugs Aging* 2010, 27: 181–192.
- [51] Wang X, Su B, Zheng L, Perry G, Smith MA, Zhu X. The role of abnormal mitochondrial dynamics in the pathogenesis of Alzheimer's disease. *J Neurochem* 2009, 109 Suppl 1: 153–159.
- [52] Wang X, Su B, Fujioka H, Zhu X. Dynamin-like protein 1 reduction underlies mitochondrial morphology and distribution abnormalities in fibroblasts from sporadic Alzheimer's disease patients. *Am J Pathol* 2008, 173: 470–482.
- [53] Chen H, McCaffery JM, Chan DC. Mitochondrial fusion protects against neurodegeneration in the cerebellum. *Cell* 2007, 130: 548–562.
- [54] Twig G, Elorza A, Molina AJ, Mohamed H, Wikstrom JD, Walzer G, *et al.* Fission and selective fusion govern mitochondrial segregation and elimination by autophagy. *EMBO J* 2008, 27: 433–446.
- [55] Wang X, Su B, Siedlak SL, Moreira PI, Fujioka H, Wang Y, *et al.* Amyloid-beta overproduction causes abnormal mitochondrial dynamics via differential modulation of mitochondrial fission/fusion proteins. *Proc Natl Acad Sci U S A* 2008, 105: 19318–19323.
- [56] Wang X, Su B, Lee HG, Li X, Perry G, Smith MA, *et al.* Impaired balance of mitochondrial fission and fusion in Alzheimer's disease. *J Neurosci* 2009, 29: 9090–9103.
- [57] Manczak M, Calkins MJ, Reddy PH. Impaired mitochondrial

- dynamics and abnormal interaction of amyloid beta with mitochondrial protein Drp1 in neurons from patients with Alzheimer's disease: implications for neuronal damage. *Hum Mol Genet* 2011, 20: 2495–2509.
- [58] Reddy PH, Tripathi R, Troung Q, Tirumala K, Reddy TP, Anekonda V, *et al.* Abnormal mitochondrial dynamics and synaptic degeneration as early events in Alzheimer's disease: implications to mitochondria-targeted antioxidant therapeutics. *Biochimica et Biophysica Acta* 2012, 1822: 639–649.
- [59] Nunomura A, Perry G, Pappolla MA, Wade R, Hirai K, Chiba S, *et al.* RNA oxidation is a prominent feature of vulnerable neurons in Alzheimer's disease. *J Neurosci* 1999, 19: 1959–1964.
- [60] Nunomura A, Tamaoki T, Motohashi N, Nakamura M, McKeel DW, Jr., Tabaton M, *et al.* The earliest stage of cognitive impairment in transition from normal aging to Alzheimer disease is marked by prominent RNA oxidation in vulnerable neurons. *Journal of Neuropathology and Experimental Neurology* 2012, 71: 233–241.
- [61] Sultana R, Baglioni M, Cecchetti R, Cai J, Klein JB, Bastiani P, *et al.* Lymphocyte mitochondria: toward identification of peripheral biomarkers in the progression of Alzheimer disease. *Free Radic Biol Med* 2013, 65: 595–606.
- [62] Smith MA, Zhu X, Tabaton M, Liu G, McKeel DW, Jr., Cohen ML, *et al.* Increased iron and free radical generation in preclinical Alzheimer disease and mild cognitive impairment. *J Alzheimers Dis* 2010, 19: 363–372.
- [63] Barone E, Di Domenico F, Sultana R, Coccia R, Mancuso C, Perluigi M, *et al.* Heme oxygenase-1 posttranslational modifications in the brain of subjects with Alzheimer disease and mild cognitive impairment. *Free Radic Biol Med* 2012, 52: 2292–2301.
- [64] Smith MA, Hirai K, Hsiao K, Pappolla MA, Harris PL, Siedlak SL, *et al.* Amyloid-beta deposition in Alzheimer transgenic mice is associated with oxidative stress. *J Neurochem* 1998, 70: 2212–2215.
- [65] Blass JP. The mitochondrial spiral. An adequate cause of dementia in the Alzheimer's syndrome. *Ann N Y Acad Sci* 2000, 924: 170–183.
- [66] Bubber P, Haroutunian V, Fisch G, Blass JP, Gibson GE. Mitochondrial abnormalities in Alzheimer brain: mechanistic implications. *Ann Neurol* 2005, 57: 695–703.
- [67] Bush AI. The metal theory of Alzheimer's disease. *J Alzheimers Dis* 2013, 33 Suppl 1: S277–281.
- [68] Smith MA, Rottkamp CA, Nunomura A, Raina AK, Perry G. Oxidative stress in Alzheimer's disease. *Biochim Biophys Acta* 2000, 1502: 139–144.
- [69] Cho HH, Cahill CM, Vanderburg CR, Scherzer CR, Wang B, Huang X, *et al.* Selective translational control of the Alzheimer amyloid precursor protein transcript by iron regulatory protein-1. *J Biol Chem* 2010, 285: 31217–31232.
- [70] Duce JA, Tsatsanis A, Cater MA, James SA, Robb E, Wikke K, *et al.* Iron-export ferroxidase activity of beta-amyloid precursor protein is inhibited by zinc in Alzheimer's disease. *Cell* 2010, 142: 857–867.
- [71] Giaccone G, Pedrotti B, Migheli A, Verga L, Perez J, Racagni G, *et al.* beta PP and Tau interaction. A possible link between amyloid and neurofibrillary tangles in Alzheimer's disease. *Am J Pathol* 1996, 148: 79–87.
- [72] Christen Y. Oxidative stress and Alzheimer disease. *Am J Clin Nutr* 2000, 71: 621S–629S.
- [73] Nunomura A, Chiba S, Lippa CF, Cras P, Kalaria RN, Takeda A, *et al.* Neuronal RNA oxidation is a prominent feature of familial Alzheimer's disease. *Neurobiol Dis* 2004, 17: 108–113.
- [74] Bogdanovic N, Zilmer M, Zilmer K, Rehema A, Karelson E. The Swedish APP670/671 Alzheimer's disease mutation: the first evidence for strikingly increased oxidative injury in the temporal inferior cortex. *Dement Geriatr Cogn Disord* 2001, 12: 364–370.
- [75] Combs CK, Karlo JC, Kao SC, Landreth GE. beta-Amyloid stimulation of microglia and monocytes results in TNFalpha-dependent expression of inducible nitric oxide synthase and neuronal apoptosis. *J Neurosci* 2001, 21: 1179–1188.
- [76] Bonda DJ, Mailankot M, Stone JG, Garrett MR, Staniszewska M, Castellani RJ, *et al.* Indoleamine 2,3-dioxygenase and 3-hydroxykynurenine modifications are found in the neuropathology of Alzheimer's disease. *Redox Rep* 2010, 15: 161–168.
- [77] Eastman CL, Guilarte TR. The role of hydrogen peroxide in the in vitro cytotoxicity of 3-hydroxykynurenine. *Neurochem Res* 1990, 15: 1101–1107.
- [78] Klivenyi P, Toldi J, Vecsei L. Kynurenines in neurodegenerative disorders: therapeutic consideration. *Adv Exp Med Biol* 2004, 541: 169–183.
- [79] Mark RJ, Lovell MA, Markesbery WR, Uchida K, Mattson MP. A role for 4-hydroxynonenal, an aldehydic product of lipid peroxidation, in disruption of ion homeostasis and neuronal death induced by amyloid beta-peptide. *J Neurochem* 1997, 68: 255–264.
- [80] Mark RJ, Pang Z, Geddes JW, Uchida K, Mattson MP. Amyloid beta-peptide impairs glucose transport in hippocampal and cortical neurons: involvement of membrane lipid peroxidation. *J Neurosci* 1997, 17: 1046–1054.
- [81] Markesbery WR, Lovell MA. Four-hydroxynonenal, a product of lipid peroxidation, is increased in the brain in Alzheimer's disease. *Neurobiol Aging* 1998, 19: 33–36.
- [82] Sayre LM, Zelasko DA, Harris PL, Perry G, Salomon RG, Smith MA. 4-Hydroxynonenal-derived advanced lipid peroxidation end products are increased in Alzheimer's

- disease. *J Neurochem* 1997, 68: 2092–2097.
- [83] Butterfield DA, Drake J, Pocernich C, Castegna A. Evidence of oxidative damage in Alzheimer's disease brain: central role for amyloid beta-peptide. *Trends Mol Med* 2001, 7: 548–554.
- [84] Smith MA, Casadesus G, Joseph JA, Perry G. Amyloid-beta and tau serve antioxidant functions in the aging and Alzheimer brain. *Free Radic Biol Med* 2002, 33: 1194–1199.
- [85] Yan SD, Yan SF, Chen X, Fu J, Chen M, Kuppusamy P, *et al.* Non-enzymatically glycosylated tau in Alzheimer's disease induces neuronal oxidant stress resulting in cytokine gene expression and release of amyloid beta-peptide. *Nat Med* 1995, 1: 693–699.
- [86] Nunomura A, Perry G, Pappolla MA, Friedland RP, Hirai K, Chiba S, *et al.* Neuronal oxidative stress precedes amyloid-beta deposition in Down syndrome. *J Neuropathol Exp Neurol* 2000, 59: 1011–1017.
- [87] Nunomura A, Perry G, Aliev G, Hirai K, Takeda A, Balraj EK, *et al.* Oxidative damage is the earliest event in Alzheimer disease. *J Neuropathol Exp Neurol* 2001, 60: 759–767.
- [88] Gomez-Isla T, Hollister R, West H, Mui S, Growdon JH, Petersen RC, *et al.* Neuronal loss correlates with but exceeds neurofibrillary tangles in Alzheimer's disease. *Ann Neurol* 1997, 41: 17–24.
- [89] Kril JJ, Patel S, Harding AJ, Halliday GM. Neuron loss from the hippocampus of Alzheimer's disease exceeds extracellular neurofibrillary tangle formation. *Acta Neuropathol (Berl)* 2002, 103: 370–376.
- [90] Kuchibhotla KV, Wegmann S, Kopeikina KJ, Hawkes J, Rudinskiy N, Andermann ML, *et al.* Neurofibrillary tangle-bearing neurons are functionally integrated in cortical circuits in vivo. *Proc Natl Acad Sci U S A* 2014, 111: 510–514.
- [91] Morsch R, Simon W, Coleman PD. Neurons may live for decades with neurofibrillary tangles. *J Neuropathol Exp Neurol* 1999, 58: 188–197.
- [92] Li HL, Wang HH, Liu SJ, Deng YQ, Zhang YJ, Tian Q, *et al.* Phosphorylation of tau antagonizes apoptosis by stabilizing beta-catenin, a mechanism involved in Alzheimer's neurodegeneration. *Proc Natl Acad Sci U S A* 2007, 104: 3591–3596.
- [93] Lin MT, Beal MF. Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* 2006, 443: 787–795.
- [94] Sherer TB, Greenamyre JT. Oxidative damage in Parkinson's disease. *Antioxid Redox Signal* 2005, 7: 627–629.
- [95] Wang X, Michaelis EK. Selective neuronal vulnerability to oxidative stress in the brain. *Front Aging Neurosci* 2010, 2: 12.
- [96] Lipski J, Nistico R, Berretta N, Guatteo E, Bernardi G, Mercuri NB. L-DOPA: a scapegoat for accelerated neurodegeneration in Parkinson's disease? *Prog Neurobiol* 2011, 94: 389–407.
- [97] Rottkamp CA, Raina AK, Zhu X, Gaier E, Bush AI, Atwood CS, *et al.* Redox-active iron mediates amyloid-beta toxicity. *Free Radic Biol Med* 2001, 30: 447–450.
- [98] Petersen RC, Thomas RG, Grundman M, Bennett D, Doody R, Ferris S, *et al.* Vitamin E and donepezil for the treatment of mild cognitive impairment. *N Engl J Med* 2005, 352: 2379–2388.
- [99] Smith RA, Kelso GF, James AM, Murphy MP. Targeting coenzyme Q derivatives to mitochondria. *Methods Enzymol* 2004, 382: 45–67.
- [100] Lu C, Zhang D, Whiteman M, Armstrong JS. Is antioxidant potential of the mitochondrial targeted ubiquinone derivative MitoQ conserved in cells lacking mtDNA? *Antioxid Redox Signal* 2008, 10: 651–660.
- [101] Murphy MP, Smith RA. Targeting antioxidants to mitochondria by conjugation to lipophilic cations. *Annu Rev Pharmacol Toxicol* 2007, 47: 629–656.
- [102] Tauskela JS. MitoQ--a mitochondria-targeted antioxidant. *IDrugs* 2007, 10: 399–412.
- [103] Siedlak SL, Casadesus G, Webber KM, Pappolla MA, Atwood CS, Smith MA, *et al.* Chronic antioxidant therapy reduces oxidative stress in a mouse model of Alzheimer's disease. *Free Radic Res* 2009, 43: 156–164.
- [104] Liu J, Head E, Gharib AM, Yuan W, Ingersoll RT, Hagen TM, *et al.* Memory loss in old rats is associated with brain mitochondrial decay and RNA/DNA oxidation: partial reversal by feeding acetyl-L-carnitine and/or R-alpha -lipoic acid. *Proc Natl Acad Sci U S A* 2002, 99: 2356–2361.
- [105] Liu J, Atamna H, Kuratsune H, Ames BN. Delaying brain mitochondrial decay and aging with mitochondrial antioxidants and metabolites. *Ann N Y Acad Sci* 2002, 959: 133–166.
- [106] Long J, Gao F, Tong L, Cotman CW, Ames BN, Liu J. Mitochondrial decay in the brains of old rats: ameliorating effect of alpha-lipoic acid and acetyl-L-carnitine. *Neurochem Res* 2009, 34: 755–763.
- [107] Aliev G, Liu J, Shenk JC, Fischbach K, Pacheco GJ, Chen SG, *et al.* Neuronal mitochondrial amelioration by feeding acetyl-L-carnitine and lipoic acid to aged rats. *J Cell Mol Med* 2009, 13(2): 320–333.
- [108] Liu J, Killilea DW, Ames BN. Age-associated mitochondrial oxidative decay: improvement of carnitine acetyltransferase substrate-binding affinity and activity in brain by feeding old rats acetyl-L-carnitine and/or R-alpha -lipoic acid. *Proc Natl Acad Sci U S A* 2002, 99: 1876–1881.
- [109] Ames BN, Liu J. Delaying the mitochondrial decay of aging with acetylcarnitine. *Ann N Y Acad Sci* 2004, 1033: 108–116.
- [110] Milgram NW, Araujo JA, Hagen TM, Treadwell BV, Ames BN. Acetyl-L-carnitine and alpha-lipoic acid supplementation of aged beagle dogs improves learning in two landmark discrimination tests. *FASEB J* 2007, 21: 3756–3762.