

## Role of autophagy in the pathogenesis of multiple sclerosis

Peizhou Liang<sup>1,2</sup>, Weidong Le<sup>2</sup>

<sup>1</sup>*Institute of Neuroscience, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Shanghai 200031, China*

<sup>2</sup>*Center for Translational Research on Neurological Diseases, the First Affiliated Hospital, Dalian Medical University, Dalian 116011, China*

Corresponding author: Weidong Le. E-mail: [wle@sibs.ac.cn](mailto:wle@sibs.ac.cn)

© Shanghai Institutes for Biological Sciences, CAS and Springer-Verlag Berlin Heidelberg 2015

Autophagy plays an important role in maintaining the cellular homeostasis. One of its functions is to degrade unnecessary organelles and proteins for energy recycling or amino-acids for cell survival. Ablation of autophagy leads to neurodegeneration. Multiple sclerosis (MS), a permanent neurological impairment typical of chronic inflammatory demyelinating disorder, is an auto-immune disease of the central nervous system (CNS). Autophagy is tightly linked to the innate and adaptive immune systems during the autoimmune process, and several studies have shown that autophagy directly participates in the progress of MS or experimental autoimmune encephalomyelitis (EAE, a mouse model of MS). Dysfunction of mitochondria that intensively influences the autophagy pathway is one of the important factors in the pathogenesis of MS. Autophagy-related gene (ATG) 5 and immune-related GTPase M (IRGM) 1 are increased, while ATG16L2 is decreased, in T-cells in EAE and active relapsing-remitting MS brains. Administration of rapamycin, an inhibitor of mammalian target of rapamycin (mTOR), ameliorates relapsing-remitting EAE. Inflammation and oxidative stress are increased in MS lesions and EAE, but Lamp2 and the LC3-II/LC3-I ratio are decreased. Furthermore, autophagy in various glial cells plays important roles in regulating neuro-inflammation in the CNS, implying potential roles in MS. In this review, we discuss the role of autophagy in the peripheral immune system and the CNS in neuro-inflammation associated with the pathogenesis of MS.

**Keywords:** autophagy; multiple sclerosis; neuro-inflammation

### Introduction

Autophagy, a lysosome-dependent degradation pathway, contributes to maintaining cellular homeostasis. Clearance of long-lived proteins, unnecessary organelles, and aggregate-prone proteins is mainly executed by autophagy for recycling cellular materials<sup>[1]</sup>. There are three subtypes of autophagy, macroautophagy, chaperone-mediated autophagy (CMA), and mitophagy. Macroautophagy is the major type for selective degradation of protein aggregates or misfolded proteins. The ubiquitin-labeled proteins are recognized by autophagy receptors, such as p62, neighbor of BRCA1 gene 1 (NBR1), and NIP3-like protein X (NIX),

to form autophagosomes that then fuse with lysosomes for degradation. Many autophagy-related genes function during this process, such as Unc51-like kinase (ULK1) and autophagy-related gene (ATG) 5. Mammalian target of rapamycin (mTOR) represses autophagy by regulating ULK1 phosphorylation, while AMPK activates this process. CMA mediates the degradation of large molecular-weight proteins containing the KFERQ motif recognized by heat shock cognate protein of 70 kDa (HSC70). The substrate is then escorted to lysosome-associated membrane protein 2 (LAMP2A) for lysosomal degradation. Mitophagy is responsible for the degradation of damaged mitochondria. Parkin and PINK1 play critical roles in this process<sup>[2]</sup>. More

attention has been paid to autophagy in the pathogenesis of neurodegenerative diseases, such as Alzheimer's disease (AD)<sup>[3]</sup>, Parkinson's disease (PD)<sup>[4]</sup> and amyotrophic lateral sclerosis (ALS)<sup>[5]</sup>.

Multiple sclerosis (MS) is a non-inherited neurodegenerative disease characterized by the involvement of innate and adaptive immunity. MS occurs mainly in young adults and its incidence in females is two to three times higher than that in males. About 85% of MS patients show relapse-remission before irreversible progression. It is widely accepted that MS is an auto-immune disease caused by autoreactive T-cells that target the myelin sheath in the central nervous system (CNS)<sup>[6, 7]</sup>. These myelin sheath-targeting T-cells infiltrate the CNS *via* breakdown of the blood brain barrier, attack the myelin sheath, and initiate chronic inflammatory responses, leading to the loss of axons and neurons. The fact that adoptive transfer of activated myelin-specific CD4<sup>+</sup> T-cells can induce experimental autoimmune encephalomyelitis (EAE) demonstrates that auto-immunity is indispensable in the pathogenesis of MS<sup>[8]</sup>. Genome-wide association studies have shown that immunologically-relevant genes, especially T-helper-cell differentiation genes, are significantly overrepresented in the pathogenesis of MS<sup>[9]</sup>. However, it is still unclear what exactly causes the disease. One of the initiators of MS is antigen from virus or bacteria that is similar to the myelin basic protein peptide<sup>[10, 11]</sup>. The auto-immunity hypothesis is now being challenged since the finding that, like many other neurodegenerative disorders, gray matter lesions and brain atrophy are detectable before MS onset<sup>[12, 13]</sup>. So the initiators of MS still need further investigation. An important factor in MS development is neuro-inflammation, which is also one of the main characteristics of MS (chronic inflammation, infiltration of peripheral immune cells, demyelination, transected axons, and gliosis). The frail neurons die when they are exposed to the inflammatory environment. Pro-inflammatory factors such as IL17 impair the differentiation of oligodendrocyte progenitor cells (OPCs) in MS<sup>[14]</sup>.

Autophagy is tightly linked with auto-immune diseases, including MS<sup>[15–17]</sup>. The extracellular matrix molecule fibronectin aggregates in chronically demyelinating MS and the relapsing phase of EAE; it may be resistant to degradation and affect the remyelination process<sup>[18]</sup>. Soluble oligomers have been detected in the brain and

cerebral spinal fluid of MS patients<sup>[19]</sup>. Meanwhile, oxidized protein aggregates increase in EAE, probably due to the reduced autophagy level<sup>[20]</sup>. Moreover, autophagy intensively regulates inflammation in various diseases. To unravel the potential role of autophagy in the pathogenesis of MS, we highlight its possible importance in both the generation of auto-reactive lymphocytes and the regulation of neuro-inflammation. Differential cell-type specificity of autophagy in the peripheral immune system and the CNS might modulate the onset and progression of MS (Fig. 1).

### Autophagy and Mitochondria in MS

Mitochondria play important roles in general and selective autophagic flux, including mitophagy. Dysfunction of mitochondria during stress is one of the important factors involved in the pathogenesis of MS and EAE. Autophagy maintains the architecture of mitochondria, restores their function, and prevents their dysfunction, which are critical for cell survival and the pathogenesis of MS<sup>[21–23]</sup>. In MS, the decreased expression of cytochrome c oxidase subunit 5b may impair the function of mitochondria<sup>[24]</sup>. Dysfunction of mitochondria produces reactive oxygen species (ROS), which contribute to demyelination and axonal loss<sup>[25]</sup>. Autophagy clears depolarized mitochondria to reduce the excessive production of ROS by increasing BECN1 or regulating ATG4 activity<sup>[26, 27]</sup>, which is protective in MS.

### Autophagy and MS in the Peripheral Immune System

#### ***Mutual Regulation of Autophagy and Inflammation***

The connections between autophagy and inflammation are complex. Each regulates the other by different mechanisms. Pattern-recognition receptors for pathogen recognition, such as toll-like receptors (TLRs) and NOD-like receptors (NLRs) can elicit autophagy for pathogen clearance<sup>[28]</sup>. However, autophagy negatively regulates inflammation to prevent the harmful amplification of inflammatory factors. For instance, ATG9 suppresses the stimulator of IFN genes protein (STING) in type-I IFN signaling to reduce the inflammatory response<sup>[29]</sup>. Blockade of autophagy leads to the activation of inflammasomes which control the proteolytic processing and secretion of IL-1 $\beta$  and IL-18 under inflammatory stress<sup>[30, 31]</sup>. Also, ablation of autophagy-related 16-like 1 (ATG16L1) increases the production of

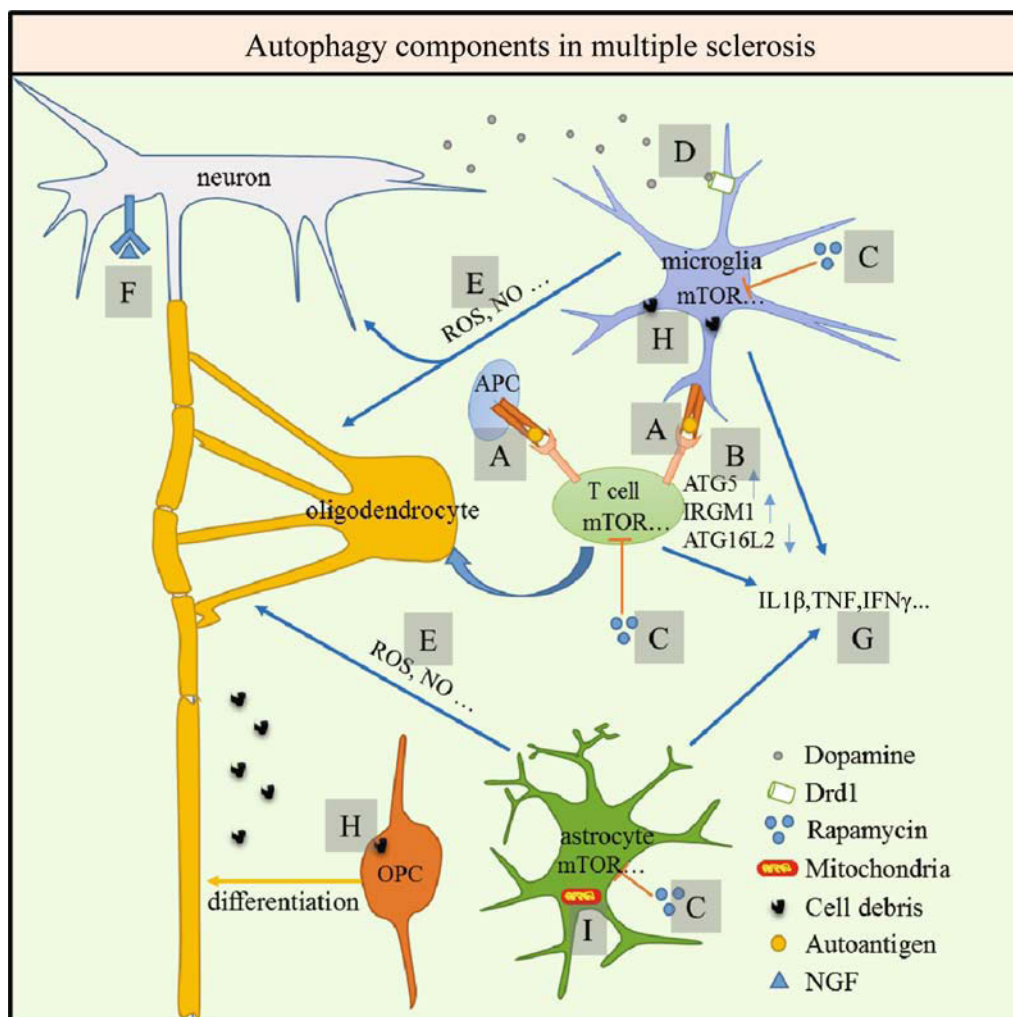


Fig. 1. Autophagy in the pathogenesis of multiple sclerosis. A: Autophagy is indispensable for antigen presentation in antigen presenting cells (APC, e.g. thymic epithelial cells, microglia). B: In MS patients and EAE mice, ATG5 and IRGM1 increase in T-cells to enhance their survival. Downregulation of ATG16L2 may perturb the homeostasis of T-cells. C: Rapamycin, the inhibitor of mTOR that represses autophagy, regulates inflammation and benefits EAE. D: Drd1 in microglia or macrophages activated by dopamine triggers autophagy to inhibit inflammasome activation during neuro-inflammation. E: ROS and NO produced by microglia and astrocytes in neuro-inflammation regulate autophagy in various cell types, including astrocytes. F: In MS patients, NGF increases in cerebrospinal fluid and represses autophagy in neurons to prevent cell death. G: Pro-inflammatory factors, such as IL-1 $\beta$ , TNF, and IFN $\gamma$ , can induce autophagy in macrophages or microglia. H: Cell debris or protein aggregates in MS or EAE are removed by microglia or macrophages through phagocytosis or autophagy. OPCs may also clear cellular debris and protein aggregates by endocytosis and autophagy. I: Autophagy is necessary to maintain the architecture and restore the function of mitochondria in astrocytes during neuro-inflammation. Depolarized mitochondria are also removed by autophagy.

IL-1 $\beta$  and IL-18 in the mouse model of Crohn's disease<sup>[32]</sup>. Moreover, infection of ATG5-deficient myeloid cells with *M. tuberculosis* increases the expression of IL-17, a potent mediator in autoimmunity<sup>[33]</sup>. In primary effective T-cells, p62 serves as a receptor for autophagic degradation of ubiquitinated BCL10 during TCR activation to protect cells

from excessively activated NF- $\kappa$ B pathway<sup>[34]</sup>.

Inflammatory factors in turn regulate the progress of autophagy. The T-helper 1 (Th1) cell cytokine IFN $\gamma$  induces autophagy in macrophages to suppress the survival of mycobacteria<sup>[35]</sup>. Another pro-inflammatory factor, Tnf, also induces autophagy in monocytes/macrophages<sup>[36]</sup>.

However, the Th2 cell cytokines IL-4 and IL-13 ablate autophagy in macrophages<sup>[36]</sup>.

### **Autoreactive T-Cell Regulation by Autophagy**

Besides innate immunity, autophagy also participates intensively in shaping the adaptive immune system. Th1 cells are crucial at the early stage of MS pathogenesis, and Th17 cells are indispensable in the later stage. Autophagy is necessary for the generation of autoreactive T-cells. It participates in multiple functions of the adaptive immune system, such as antigen presentation and maintenance of the homeostasis of T-cells.

Autophagy mediates antigen-presentation during the generation of autoreactive T-cells. Autophagy enhances antigen-presentation to CD4<sup>+</sup> T-cells via major histocompatibility complex class II molecules<sup>[37, 38]</sup>. In thymic epithelial cells, autophagy contributes to the positive and negative control of the T-cell repertoire, playing a critical role in the elimination of self-reactive T-cells and exotic pathogens. Mice thymi with ATG5 ablation show infiltration of autoreactive CD4<sup>+</sup> T-cells into multiple organs and induce autoimmune colitis similar to Crohn's disease-related phenotypes<sup>[38]</sup>. Atg7 ablation in dendritic cells (DCs), the most potent antigen-presenting cells, ameliorates EAE. Blockade of autophagy in DCs by chloroquine also delays the onset or reduces the severity of EAE<sup>[39]</sup>. DCs from patients with Crohn's disease with ATG16L1 and NOD2 variants are defective in antigen presentation<sup>[40]</sup>. The presentation of citrullinated peptides by DCs, macrophages, and thymic DCs is blocked by inhibition of autophagy with 3-methyladenine<sup>[41]</sup>.

Autophagy regulates the homeostasis, proliferation, differentiation, maturation, and apoptosis of T-cells. It also maintains the calcium flux and the pro-survival process and counteracts pro-apoptotic action to maintain T-cell homeostasis<sup>[42–45]</sup>. Cyt1 induces autophagosome formation when it binds to CD46, which may function in maintaining the homeostasis of T-cells during adaptive immunity<sup>[46]</sup>. Moreover, the expression of ATG16L2 in T-cells is reduced, and this is likely to perturb the homeostasis of T-cells in MS<sup>[47]</sup>. ATG5 plays important roles in T-cell survival and controls the proliferation, differentiation and maturation of CD4/8<sup>+</sup> T-cells and B-cells<sup>[48–50]</sup>. mTOR-deficient T-cells fail to differentiate into T-helper cells, Th1, Th2, and Th17 effector cells<sup>[51, 52]</sup>. ATG5 is significantly elevated in T-cells

in MS lesions and EAE mice compared to controls. The increase of autophagy in T-cells may promote their survival, contributing to the pathogenesis of MS<sup>[53]</sup>. Immune-related GTPase M1 is highly expressed in MS lesions in the CNS and EAE models, and contributes to the pathogenesis of MS by increasing the survival of autoreactive CD4<sup>+</sup> T-cells<sup>[17]</sup>. Rapamycin, an inhibitor of mTOR, ameliorates the relapsing-remitting EAE (RR-EAE) in SJL/j mice by increasing the Treg cell population<sup>[54]</sup>. However, the contributions of autophagy in different T-cell populations to cell death depend on the context. Upon activation of autophagy, Th2 cells undergo growth factor-withdrawal-mediated cell death<sup>[55]</sup>. The induction of autophagy through binding of HIV-1 envelope glycoproteins to CXCR4 chemokine receptor 4 of CD4<sup>+</sup> T-cells is necessary for apoptosis<sup>[56]</sup>. FADD or casp8 in T-cells limits autophagy and protects against cell necrosis and inflammation<sup>[57, 58]</sup>. Alteration of the proportions of different T-cell groups affects the immune response<sup>[59]</sup>.

## **Pathological Role of Autophagy in MS**

### **Autophagy and Neuro-inflammation**

CNS autophagy is linked with serious neurodegenerative diseases. Deficiency of basal autophagy in neurons leads to protein aggregation and finally neurodegenerative diseases<sup>[60, 61]</sup>. Neuro-inflammation is one of the important factors that cause neurodegenerative diseases, especially MS. Autophagy and neuro-inflammation regulate each other to affect the progression of various diseases in the CNS.

Neuro-inflammation is negatively regulated by autophagy to avoid its harm to the CNS. Microglia, resident immune cells in the CNS, play important roles in the initiation and sustenance of neuro-inflammation to clear pathogens. They produce pro-inflammatory factors and neurotoxic factors including ROS and nitric oxide (NO). Subsequently-activated astrocytes further amplify the inflammatory response to produce more pro-inflammatory factors, chemokines, and NO, which have toxic effects on primary neurons<sup>[7, 62]</sup>. However, autophagy plays an essential role in maintaining neuro-inflammation at a harmless level. Activation of Drd1 by dopamine inhibits NOD-LRR-containing pyrin domain 3 (NLRP3) inflammasome activation through autophagic degradation



of NLRP3 *via* the E3 ubiquitin ligase MARCH7<sup>[63]</sup>. The dopamine system also regulates lymphocyte activity in lupus and MS, suggesting an important role of dopamine in the regulation of peripheral or CNS infiltrated lymphocytes during the pathogenesis of MS<sup>[64, 65]</sup>. In addition, inhibition of neuro-inflammation by autophagy has great benefits on ischemia<sup>[66]</sup>. Autophagy may be a stress response to the negative feedback regulation of neuro-inflammation. In the chronic LPS model, pro-inflammatory factor IL-1 $\beta$  in cortex is positively correlated with the autophagy markers Beclin-1 and LC3-II and inversely correlated with p62<sup>[67]</sup>.

Autophagy can be deleterious depending on the disease. Activation of the autophagic pathway in ischemia leads to the death of astrocytes and neurons, which can be rescued by autophagy inhibitors<sup>[68, 69]</sup>. Meanwhile, blockade of microglial autophagy in permanent middle cerebral artery occlusion by autophagy inhibitors reduces the severity of the disease<sup>[70]</sup>. Thus, autophagy in different diseases and cell types in the CNS seems to function differently. Autophagic cell death is still under debate and needs further investigation<sup>[71]</sup>.

### **Autophagy in Glial Cells**

Until recently, most studies on autophagy in the CNS focused on neurons, with less on glial cells. The activation of glial cells plays important roles in neuro-inflammation, and these cells are indispensable for the development of MS<sup>[7]</sup>. MS progression occurs between glial activation and gliosis. Autophagy in various glial cells may play a role in responding to stress in MS. Therefore, it is urgent to study the effect of glial autophagy on neurological diseases, including MS.

Microglia are the first line of defense in the CNS, monitoring exotic and intrinsic pathogens. During demyelination, cell debris is mainly phagocytized by microglia or macrophages, a process that requires autophagy-related genes<sup>[72]</sup>. Impairment of autophagy in microglia may hinder debris clearance, leading to damaged remyelination and augmentation of persistent neuro-inflammation. The rapid activation of microglia produces pro-inflammatory cytokines and free radicals, and acts as the major antigen-presenting cell<sup>[73, 74]</sup>. Pro-inflammatory factors such as IL-1 $\beta$  can trigger microglial autophagy, suggesting links between autophagy and inflammation activation<sup>[75]</sup>. mTOR plays a pivotal role in autophagy suppression and promotes inflammation in microglia, and

rapamycin may have a beneficial effect in the treatment of MS<sup>[76]</sup>. mTOR inhibitors reduce neuro-inflammation by inhibiting microglial activation or viability, decreasing pro-inflammatory cytokines<sup>[77]</sup>. Simian immunodeficiency virus-infected microglia attenuate the survival of neurons by inhibiting neuronal autophagy<sup>[78]</sup>, which implicates a cell-type regulation of autophagy in the CNS. However, microglial autophagy in middle cerebral artery occlusion makes the disease worse<sup>[70]</sup>. It is possible that the role of microglial autophagy depends on the disease.

Astrocytes are the primary supportive elements for neuronal structure and neurotrophs, comprising ~50% of cells in the adult mammalian brain. Autophagy is necessary for the differentiation and maturation of astrocytes<sup>[79]</sup>, and can be initiated in inflammatory stress and may be important in the development of MS. During pro-inflammatory stimuli, autophagy is induced to maintain mitochondrial networks<sup>[22]</sup>. Impairment of the autophagy pathway by ablation of Atg7 in astrocytes in the inflammatory environment amplifies the response to produce more neurotoxic factors such as ROS, which induces astrocytic cell death<sup>[22, 80]</sup>. mTOR in the astrocyte may play a protective role in ischemia *via* its downstream kinase S6K1<sup>[81]</sup>. mTOR can also regulate the stability of iNOS mRNA in astrocytes, reducing the neurotoxic effect of NO which impairs autophagy by disrupting the BECN1 complex and activates mTORC1<sup>[82-84]</sup>.

OPCs are indispensable during oligodendrocyte differentiation and myelin remyelination; they are activated for remyelination in MS lesions<sup>[85, 86]</sup>. They also play an important role in MS pathogenesis by modulating immune activity in the CNS. Act1 deletion in OPCs blocks the IL-17 pathway, attenuating neuro-inflammation and increasing OPC maturation<sup>[14]</sup>. OPCs can also clear  $\beta$ -amyloid peptides by triggering endocytosis and autophagy<sup>[87]</sup>, implying an autophagic role of OPCs in MS. However, the exact functions of OPCs in MS need further investigation, including those of autophagy and its differentiation, nutritional support, debris clearance, and inflammation regulation.

### **Roles of Autophagy in MS-Associated Demyelination and Remyelination**

Autophagy is closely linked to demyelination and remyelination. It plays potential roles in improving

Schwann cell remyelination in demyelinating peripheral neuropathies<sup>[88]</sup>. Rapamycin, a potent autophagy inducer, improves myelination, leading to enhanced neuronal survival in tuberous sclerosis<sup>[89]</sup>. Meanwhile, in Long–Evans shaker rats, an increased autophagy level increases the number of myelinated axons and myelin sheath thickness during dysmyelination and demyelination, which implies that the autophagy pathway is a direct target for therapy for demyelination<sup>[90]</sup>. Furthermore, high mobility group box chromosomal protein 1 (HMGB1), which promotes autophagy, is elevated in MS and EAE<sup>[91]</sup>. However, the progression of MS and EAE involves many cell types. Different cell types in the lesion site might show different patterns of autophagy. In MS, especially in the acute phase, nerve growth factor (NGF) is dramatically increased in the cerebrospinal fluid<sup>[92]</sup>. NGF inhibits autophagy and cell death of neurons through the p75 neurotrophin receptor, suggesting a protective role in MS<sup>[93]</sup>. In MS lesions, Lamp2 expression is reduced<sup>[94]</sup>, implying an impairment of autophagy. The mTOR signaling pathway that inhibits autophagy restores the regrowth of axons in the CNS, which is important for remyelination in MS<sup>[95, 96]</sup>. Moreover, in acute and chronic EAE, protein aggregates in the spinal cord and a reduced LC3-II/LC3-I ratio suggest that the protein turnover mechanism through autophagy is impaired<sup>[20]</sup>. In fact, the mechanism of autophagy in the pathogenesis of MS in the CNS is still superficially understood. Autophagy in different stages of the disease and different cell types needs further investigation to determine the causal relationship between autophagy and MS.

### Conclusive Remarks

So far, little is known about the exact mechanism of autophagy in MS. It is still controversial whether autophagy leads to cell death in MS or is a rescue mechanism activated as part of an endogenous neuroprotective response, since autophagy plays opposite functions in inflammation and cell survival, depending on the context. Drugs modifying the immune system and remyelination have been used to help with relapse management, reduce the degree of disability, and improve the quality of life<sup>[97, 98]</sup>. The effects of rapamycin and chloroquine in the therapy of EAE shed light on the autophagy pathway as a potential

target for drug development. The inhibition of autophagy in mesenchymal stem cells provides a novel strategy to improve therapeutic effects by enhancing the suppression of CD4<sup>+</sup> T-cells<sup>[99]</sup>. In order to develop effective and more specific therapeutic strategies, we need to fully understand the complex interplay between autophagy and MS in different cell types.

### ACKNOWLEDGEMENTS

This review was supported by a grant from the National Natural Science Foundation of China (81430021 and 81370470), China Postdoctoral Science Foundation (2014M551465) and the Collaborative Innovation Center for Brain Science.

Received date: 2015-05-20; Accepted date: 2015-07-05

### REFERENCES

- [1] Rubinsztein DC. The roles of intracellular protein-degradation pathways in neurodegeneration. *Nature* 2006, 443: 780–786.
- [2] Nixon RA. The role of autophagy in neurodegenerative disease. *Nat Med* 2013, 19: 983–997.
- [3] Nixon RA, Wegiel J, Kumar A, Yu WH, Peterhoff C, Cataldo A, *et al.* Extensive involvement of autophagy in Alzheimer disease: an immuno-electron microscopy study. *J Neuropathol Exp Neurol* 2005, 64: 113–122.
- [4] Lynch-Day MA, Mao K, Wang K, Zhao M, Klionsky DJ. The role of autophagy in Parkinson's disease. *Cold Spring Harb Perspect Med* 2012, 2: a009357.
- [5] Song CY, Guo JF, Liu Y, Tang BS. Autophagy and its comprehensive impact on ALS. *Int J Neurosci* 2012, 122: 695–703.
- [6] Steinman L. Multiple sclerosis: a two-stage disease. *Nat Immunol* 2001, 2: 762–764.
- [7] Glass CK, Saijo K, Winner B, Marchetto MC, Gage FH. Mechanisms underlying inflammation in neurodegeneration. *Cell* 2010, 140: 918–934.
- [8] Ben-Nun A, Wekerle H, Cohen IR. The rapid isolation of clonable antigen-specific T lymphocyte lines capable of mediating autoimmune encephalomyelitis. *Eur J Immunol* 1981, 11: 195–199.
- [9] International Multiple Sclerosis Genetics C, Wellcome Trust Case Control C, Sawcer S, Hellenthal G, Pirinen M, Spencer CC, *et al.* Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. *Nature* 2011, 476: 214–219.
- [10] Sospedra M, Martin R. Immunology of multiple sclerosis. *Annu Rev Immunol* 2005, 23: 683–747.
- [11] Fujinami RS, Oldstone MB. Amino acid homology between

- the encephalitogenic site of myelin basic protein and virus: mechanism for autoimmunity. *Science* 1985, 230: 1043–1045.
- [12] Stys PK, Zamponi GW, van Minnen J, Geurts JJ. Will the real multiple sclerosis please stand up? *Nat Rev Neurosci* 2012, 13: 507–514.
- [13] Ellwardt E, Zipp F. Molecular mechanisms linking neuroinflammation and neurodegeneration in MS. *Exp Neurol* 2014, 262 Pt A: 8–17.
- [14] Kang Z, Wang C, Zepp J, Wu L, Sun K, Zhao J, *et al.* Act1 mediates IL-17-induced EAE pathogenesis selectively in NG2(+) glial cells. *Nat Neurosci* 2013, 16: 1401–1408.
- [15] Wellcome Trust Case Control C. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature* 2007, 447: 661–678.
- [16] Zhou XJ, Lu XL, Lv JC, Yang HZ, Qin LX, Zhao MH, *et al.* Genetic association of PRDM1-ATG5 intergenic region and autophagy with systemic lupus erythematosus in a Chinese population. *Ann Rheum Dis* 2011, 70: 1330–1337.
- [17] Xu H, Wu ZY, Fang F, Guo L, Chen D, Chen JX, *et al.* Genetic deficiency of Irgm1 (LRG-47) suppresses induction of experimental autoimmune encephalomyelitis by promoting apoptosis of activated CD4+ T cells. *FASEB J* 2010, 24: 1583–1592.
- [18] Stoffels JM, de Jonge JC, Stancic M, Nomden A, van Strien ME, Ma D, *et al.* Fibronectin aggregation in multiple sclerosis lesions impairs remyelination. *Brain* 2013, 136: 116–131.
- [19] David MA, Tayebi M. Detection of protein aggregates in brain and cerebrospinal fluid derived from multiple sclerosis patients. *Front Neurol* 2014, 5: 251.
- [20] Dasgupta A, Zheng J, Perrone-Bizzozero NI, Bizzozero OA. Increased carbonylation, protein aggregation and apoptosis in the spinal cord of mice with experimental autoimmune encephalomyelitis. *ASN Neuro* 2013, 5: e00111.
- [21] Kalman B, Laitinen K, Komoly S. The involvement of mitochondria in the pathogenesis of multiple sclerosis. *J Neuroimmunol* 2007, 188: 1–12.
- [22] Motori E, Puyal J, Toni N, Ghanem A, Angeloni C, Malaguti M, *et al.* Inflammation-induced alteration of astrocyte mitochondrial dynamics requires autophagy for mitochondrial network maintenance. *Cell Metab* 2013, 18: 844–859.
- [23] Lu Q, Zhang J, Allison R, Gay H, Yang WX, Bhowmick NA, *et al.* Identification of extracellular delta-catenin accumulation for prostate cancer detection. *Prostate* 2009, 69: 411–418.
- [24] Broadwater L, Pandit A, Clements R, Azzam S, Vadnal J, Sulak M, *et al.* Analysis of the mitochondrial proteome in multiple sclerosis cortex. *Biochim Biophys Acta* 2011, 1812: 630–641.
- [25] van Horssen J, Witte ME, Schreibelt G, de Vries HE. Radical changes in multiple sclerosis pathogenesis. *Biochim Biophys Acta* 2011, 1812: 141–150.
- [26] Chen Y, Gibson SB. Is mitochondrial generation of reactive oxygen species a trigger for autophagy? *Autophagy* 2008, 4: 246–248.
- [27] Chen Y, McMillan-Ward E, Kong J, Israels SJ, Gibson SB. Oxidative stress induces autophagic cell death independent of apoptosis in transformed and cancer cells. *Cell Death Differ* 2008, 15: 171–182.
- [28] Saitoh T, Akira S. Regulation of innate immune responses by autophagy-related proteins. *J Cell Biol* 2010, 189: 925–935.
- [29] Saitoh T, Fujita N, Hayashi T, Takahara K, Satoh T, Lee H, *et al.* Atg9a controls dsDNA-driven dynamic translocation of STING and the innate immune response. *Proc Natl Acad Sci U S A* 2009, 106: 20842–20846.
- [30] Deretic V, Saitoh T, Akira S. Autophagy in infection, inflammation and immunity. *Nat Rev Immunol* 2013, 13: 722–737.
- [31] Shi CS, Shenderov K, Huang NN, Kabat J, Abu-Asab M, Fitzgerald KA, *et al.* Activation of autophagy by inflammatory signals limits IL-1beta production by targeting ubiquitinated inflammasomes for destruction. *Nat Immunol* 2012, 13: 255–263.
- [32] Saitoh T, Fujita N, Jang MH, Uematsu S, Yang BG, Satoh T, *et al.* Loss of the autophagy protein Atg16L1 enhances endotoxin-induced IL-1beta production. *Nature* 2008, 456: 264–268.
- [33] Castillo EF, Dekonenko A, Arko-Mensah J, Mandell MA, Dupont N, Jiang S, *et al.* Autophagy protects against active tuberculosis by suppressing bacterial burden and inflammation. *Proc Natl Acad Sci U S A* 2012, 109: E3168–3176.
- [34] Paul S, Kashyap AK, Jia W, He YW, Schaefer BC. Selective autophagy of the adaptor protein Bcl10 modulates T cell receptor activation of NF-kappaB. *Immunity* 2012, 36: 947–958.
- [35] Gutierrez MG, Master SS, Singh SB, Taylor GA, Colombo MI, Deretic V. Autophagy is a defense mechanism inhibiting BCG and Mycobacterium tuberculosis survival in infected macrophages. *Cell* 2004, 119: 753–766.
- [36] Harris J, Keane J. How tumour necrosis factor blockers interfere with tuberculosis immunity. *Clin Exp Immunol* 2010, 161: 1–9.
- [37] Paludan C, Schmid D, Landthaler M, Vockerodt M, Kube D, Tuschl T, *et al.* Endogenous MHC class II processing of a viral nuclear antigen after autophagy. *Science* 2005, 307: 593–596.
- [38] Nedjic J, Aichinger M, Emmerich J, Mizushima N, Klein L. Autophagy in thymic epithelium shapes the T-cell repertoire and is essential for tolerance. *Nature* 2008, 455: 396–400.
- [39] Bhattacharya A, Parillon X, Zeng S, Han S, Eissa NT.

- Deficiency of autophagy in dendritic cells protects against experimental autoimmune encephalomyelitis. *J Biol Chem* 2014, 289: 26525–26532.
- [40] Cooney R, Baker J, Brain O, Danis B, Pichulik T, Allan P, *et al.* NOD2 stimulation induces autophagy in dendritic cells influencing bacterial handling and antigen presentation. *Nat Med* 2010, 16: 90–97.
- [41] Ireland JM, Unanue ER. Autophagy in antigen-presenting cells results in presentation of citrullinated peptides to CD4 T cells. *J Exp Med* 2011, 208: 2625–2632.
- [42] Jia W, Pua HH, Li QJ, He YW. Autophagy regulates endoplasmic reticulum homeostasis and calcium mobilization in T lymphocytes. *J Immunol* 2011, 186: 1564–1574.
- [43] Lee JS, Li Q, Lee JY, Lee SH, Jeong JH, Lee HR, *et al.* FLIP-mediated autophagy regulation in cell death control. *Nat Cell Biol* 2009, 11: 1355–1362.
- [44] He MX, He YW. A role for c-FLIP(L) in the regulation of apoptosis, autophagy, and necroptosis in T lymphocytes. *Cell Death Differ* 2013, 20: 188–197.
- [45] Hubbard VM, Valdor R, Patel B, Singh R, Cuervo AM, Macian F. Macroautophagy regulates energy metabolism during effector T cell activation. *J Immunol* 2010, 185: 7349–7357.
- [46] Ni Choileain S, Astier AL. CD46 plasticity and its inflammatory bias in multiple sclerosis. *Arch Immunol Ther Exp (Warsz)* 2011, 59: 49–59.
- [47] Yin L, Liu J, Dong H, Xu E, Qiao Y, Wang L, *et al.* Autophagy-related gene16L2, a potential serum biomarker of multiple sclerosis evaluated by bead-based proteomic technology. *Neurosci Lett* 2014, 562: 34–38.
- [48] Miller BC, Zhao Z, Stephenson LM, Cadwell K, Pua HH, Lee HK, *et al.* The autophagy gene ATG5 plays an essential role in B lymphocyte development. *Autophagy* 2008, 4: 309–314.
- [49] Pua HH, He YW. Maintaining T lymphocyte homeostasis: another duty of autophagy. *Autophagy* 2007, 3: 266–267.
- [50] Pua HH, Dzhagalov I, Chuck M, Mizushima N, He YW. A critical role for the autophagy gene Atg5 in T cell survival and proliferation. *J Exp Med* 2007, 204: 25–31.
- [51] Delgoffe GM, Kole TP, Zheng Y, Zarek PE, Matthews KL, Xiao B, *et al.* The mTOR kinase differentially regulates effector and regulatory T cell lineage commitment. *Immunity* 2009, 30: 832–844.
- [52] Chi H. Regulation and function of mTOR signalling in T cell fate decisions. *Nat Rev Immunol* 2012, 12: 325–338.
- [53] Alirezai M, Fox HS, Flynn CT, Moore CS, Hebb AL, Frausto RF, *et al.* Elevated ATG5 expression in autoimmune demyelination and multiple sclerosis. *Autophagy* 2009, 5: 152–158.
- [54] Esposito M, Ruffini F, Bellone M, Gagliani N, Battaglia M, Martino G, *et al.* Rapamycin inhibits relapsing experimental autoimmune encephalomyelitis by both effector and regulatory T cells modulation. *J Neuroimmunol* 2010, 220: 52–63.
- [55] Li C, Capan E, Zhao Y, Zhao J, Stolz D, Watkins SC, *et al.* Autophagy is induced in CD4+ T cells and important for the growth factor-withdrawal cell death. *J Immunol* 2006, 177: 5163–5168.
- [56] Espert L, Denizot M, Grimaldi M, Robert-Hebmann V, Gay B, Varbanov M, *et al.* Autophagy is involved in T cell death after binding of HIV-1 envelope proteins to CXCR4. *J Clin Invest* 2006, 116: 2161–2172.
- [57] Bell BD, Leverrier S, Weist BM, Newton RH, Arechiga AF, Luhrs KA, *et al.* FADD and caspase-8 control the outcome of autophagic signaling in proliferating T cells. *Proc Natl Acad Sci U S A* 2008, 105: 16677–16682.
- [58] Yu L, Alva A, Su H, Dutt P, Freundt E, Welsh S, *et al.* Regulation of an ATG7-beclin 1 program of autophagic cell death by caspase-8. *Science* 2004, 304: 1500–1502.
- [59] Oreja-Guevara C, Ramos-Cejudo J, Aroeira LS, Chamorro B, Diez-Tejedor E. TH1/TH2 Cytokine profile in relapsing-remitting multiple sclerosis patients treated with Glatiramer acetate or Natalizumab. *BMC Neurol* 2012, 12: 95.
- [60] Komatsu M, Waguri S, Chiba T, Murata S, Iwata J, Tanida I, *et al.* Loss of autophagy in the central nervous system causes neurodegeneration in mice. *Nature* 2006, 441: 880–884.
- [61] Hara T, Nakamura K, Matsui M, Yamamoto A, Nakahara Y, Suzuki-Migishima R, *et al.* Suppression of basal autophagy in neural cells causes neurodegenerative disease in mice. *Nature* 2006, 441: 885–889.
- [62] Saijo K, Winner B, Carson CT, Collier JG, Boyer L, Rosenfeld MG, *et al.* A Nurr1/CoREST pathway in microglia and astrocytes protects dopaminergic neurons from inflammation-induced death. *Cell* 2009, 137: 47–59.
- [63] Yan Y, Jiang W, Liu L, Wang X, Ding C, Tian Z, *et al.* Dopamine controls systemic inflammation through inhibition of NLRP3 inflammasome. *Cell* 2015, 160: 62–73.
- [64] Besser MJ, Ganor Y, Levite M. Dopamine by itself activates either D2, D3 or D1/D5 dopaminergic receptors in normal human T-cells and triggers the selective secretion of either IL-10, TNFalpha or both. *J Neuroimmunol* 2005, 169: 161–171.
- [65] Pacheco R, Contreras F, Zouali M. The dopaminergic system in autoimmune diseases. *Front Immunol* 2014, 5: 117.
- [66] Zhou X, Zhou J, Li X, Guo C, Fang T, Chen Z. GSK-3beta inhibitors suppressed neuroinflammation in rat cortex by activating autophagy in ischemic brain injury. *Biochem Biophys Res Commun* 2011, 411: 271–275.
- [67] Francois A, Terro F, Quellard N, Fernandez B, Chassaing D, Janet T, *et al.* Impairment of autophagy in the central nervous system during lipopolysaccharide-induced inflammatory stress in mice. *Mol Brain* 2014, 7: 56.



- [68] Qin AP, Liu CF, Qin YY, Hong LZ, Xu M, Yang L, *et al.* Autophagy was activated in injured astrocytes and mildly decreased cell survival following glucose and oxygen deprivation and focal cerebral ischemia. *Autophagy* 2010, 6: 738–753.
- [69] Wang JY, Xia Q, Chu KT, Pan J, Sun LN, Zeng B, *et al.* Severe global cerebral ischemia-induced programmed necrosis of hippocampal CA1 neurons in rat is prevented by 3-methyladenine: a widely used inhibitor of autophagy. *J Neuropathol Exp Neurol* 2011, 70: 314–322.
- [70] Yang Z, Zhong L, Zhong S, Xian R, Yuan B. Hypoxia induces microglia autophagy and neural inflammation injury in focal cerebral ischemia model. *Exp Mol Pathol* 2015, 98: 219–224.
- [71] Ghavami S, Shojaei S, Yeganeh B, Ande SR, Jangamreddy JR, Mehrpour M, *et al.* Autophagy and apoptosis dysfunction in neurodegenerative disorders. *Prog Neurobiol* 2014, 112: 24–49.
- [72] Sanjuan MA, Dillon CP, Tait SW, Moshiah S, Dorsey F, Connell S, *et al.* Toll-like receptor signalling in macrophages links the autophagy pathway to phagocytosis. *Nature* 2007, 450: 1253–1257.
- [73] Benveniste EN. Role of macrophages/microglia in multiple sclerosis and experimental allergic encephalomyelitis. *J Mol Med (Berl)* 1997, 75: 165–173.
- [74] Chastain EM, Duncan DS, Rodgers JM, Miller SD. The role of antigen presenting cells in multiple sclerosis. *Biochim Biophys Acta* 2011, 1812: 265–274.
- [75] Francois A, Terro F, Janet T, Rioux Bilan A, Paccalin M, Page G. Involvement of interleukin-1 $\beta$  in the autophagic process of microglia: relevance to Alzheimer's disease. *J Neuroinflammation* 2013, 10: 151.
- [76] Dello Russo C, Lisi L, Feinstein DL, Navarra P. mTOR kinase, a key player in the regulation of glial functions: relevance for the therapy of multiple sclerosis. *Glia* 2013, 61: 301–311.
- [77] Dello Russo C, Lisi L, Tringali G, Navarra P. Involvement of mTOR kinase in cytokine-dependent microglial activation and cell proliferation. *Biochem Pharmacol* 2009, 78: 1242–1251.
- [78] Alirezaei M, Kiosses WB, Flynn CT, Brady NR, Fox HS. Disruption of neuronal autophagy by infected microglia results in neurodegeneration. *PLoS One* 2008, 3: e2906.
- [79] Wang S, Li B, Qiao H, Lv X, Liang Q, Shi Z, *et al.* Autophagy-related gene Atg5 is essential for astrocyte differentiation in the developing mouse cortex. *EMBO Rep* 2014, 15: 1053–1061.
- [80] Lee SJ, Cho KS, Koh JY. Oxidative injury triggers autophagy in astrocytes: the role of endogenous zinc. *Glia* 2009, 57: 1351–1361.
- [81] Pastor MD, Garcia-Yebenes I, Fradejas N, Perez-Ortiz JM, Mora-Lee S, Tranque P, *et al.* mTOR/S6 kinase pathway contributes to astrocyte survival during ischemia. *J Biol Chem* 2009, 284: 22067–22078.
- [82] Cross AH, Manning PT, Keeling RM, Schmidt RE, Misko TP. Peroxynitrite formation within the central nervous system in active multiple sclerosis. *J Neuroimmunol* 1998, 88: 45–56.
- [83] Lisi L, Navarra P, Feinstein DL, Dello Russo C. The mTOR kinase inhibitor rapamycin decreases iNOS mRNA stability in astrocytes. *J Neuroinflammation* 2011, 8: 1.
- [84] Sarkar S, Korolchuk VI, Renna M, Imarisio S, Fleming A, Williams A, *et al.* Complex inhibitory effects of nitric oxide on autophagy. *Mol Cell* 2011, 43: 19–32.
- [85] Chang A, Nishiyama A, Peterson J, Prineas J, Trapp BD. NG2-positive oligodendrocyte progenitor cells in adult human brain and multiple sclerosis lesions. *J Neurosci* 2000, 20: 6404–6412.
- [86] Wolswijk G. Oligodendrocyte precursor cells in the demyelinated multiple sclerosis spinal cord. *Brain* 2002, 125: 338–349.
- [87] Li W, Tang Y, Fan Z, Meng Y, Yang G, Luo J, *et al.* Autophagy is involved in oligodendroglial precursor-mediated clearance of amyloid peptide. *Mol Neurodegener* 2013, 8: 27.
- [88] Rangaraju S, Verrier JD, Madorsky I, Nicks J, Dunn WA, Jr., Notterpek L. Rapamycin activates autophagy and improves myelination in explant cultures from neuropathic mice. *J Neurosci* 2010, 30: 11388–11397.
- [89] Meikle L, Pollizzi K, Egnor A, Kramvis I, Lane H, Sahin M, *et al.* Response of a neuronal model of tuberous sclerosis to mammalian target of rapamycin (mTOR) inhibitors: effects on mTORC1 and Akt signaling lead to improved survival and function. *J Neurosci* 2008, 28: 5422–5432.
- [90] Smith CM, Mayer JA, Duncan ID. Autophagy promotes oligodendrocyte survival and function following dysmyelination in a long-lived myelin mutant. *J Neurosci* 2013, 33: 8088–8100.
- [91] Andersson A, Covacu R, Sunnemark D, Danilov AI, Dal Bianco A, Khademi M, *et al.* Pivotal advance: HMGB1 expression in active lesions of human and experimental multiple sclerosis. *J Leukoc Biol* 2008, 84: 1248–1255.
- [92] Laudiero LB, Aloe L, Levi-Montalcini R, Buttinelli C, Schilter D, Gillessen S, *et al.* Multiple sclerosis patients express increased levels of beta-nerve growth factor in cerebrospinal fluid. *Neurosci Lett* 1992, 147: 9–12.
- [93] Florez-McClure ML, Linseman DA, Chu CT, Barker PA, Bouchard RJ, Le SS, *et al.* The p75 neurotrophin receptor can induce autophagy and death of cerebellar Purkinje neurons. *J Neurosci* 2004, 24: 4498–4509.
- [94] Lindberg RL, De Groot CJ, Certa U, Ravid R, Hoffmann F, Kappos L, *et al.* Multiple sclerosis as a generalized CNS disease--comparative microarray analysis of normal appearing white matter and lesions in secondary progressive MS. *J Neuroimmunol* 2004, 152: 154–167.

- [95] Kim SR, Chen X, Oo TF, Kareva T, Yarygina O, Wang C, *et al.* Dopaminergic pathway reconstruction by Akt/Rheb-induced axon regeneration. *Ann Neurol* 2011, 70: 110–120.
- [96] Park KK, Liu K, Hu Y, Smith PD, Wang C, Cai B, *et al.* Promoting axon regeneration in the adult CNS by modulation of the PTEN/mTOR pathway. *Science* 2008, 322: 963–966.
- [97] Tullman MJ. A review of current and emerging therapeutic strategies in multiple sclerosis. *Am J Manag Care* 2013, 19: S21–27.
- [98] Zhang Y, Guo TB, Lu H. Promoting remyelination for the treatment of multiple sclerosis: opportunities and challenges. *Neurosci Bull* 2013, 29: 144–154.
- [99] Dang S, Xu H, Xu C, Cai W, Li Q, Cheng Y, *et al.* Autophagy regulates the therapeutic potential of mesenchymal stem cells in experimental autoimmune encephalomyelitis. *Autophagy* 2014, 10: 1301–1315.